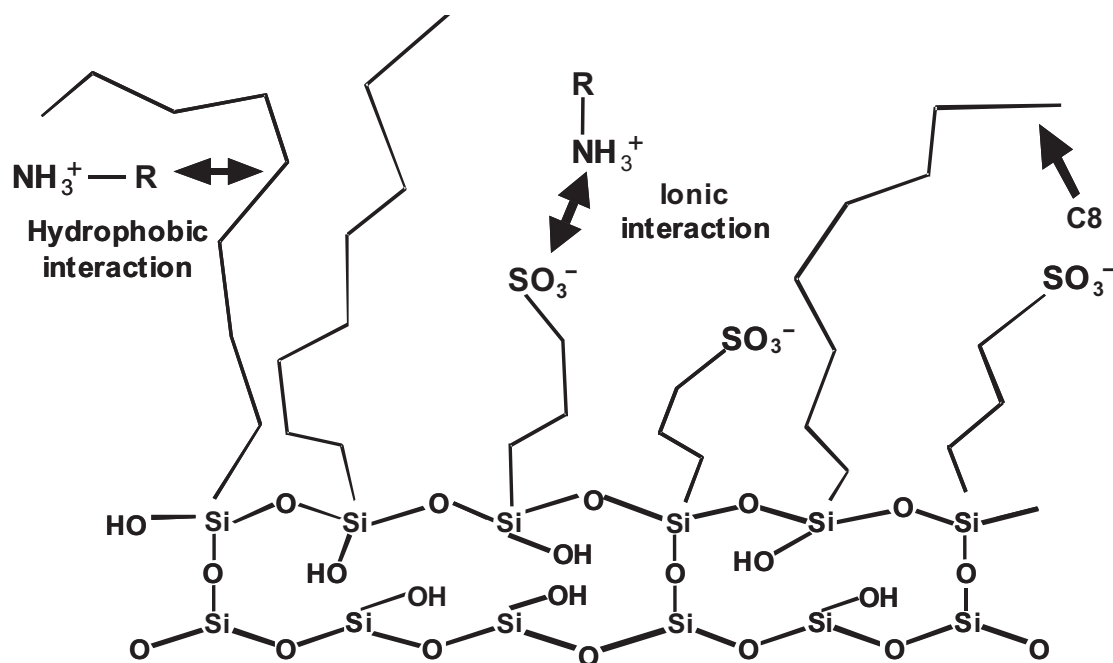


## SAMPLE PREPARATION BY MIXED-MODE SPE USING ISOLUTE® HCX

This technical note describes the extraction of basic drugs from biological fluids using ISOLUTE HCX mixed-mode SPE sorbents.

Sample preparation techniques such as protein precipitation, supported liquid extraction or non-polar SPE may not be selective enough to give extracts of sufficient purity for low level analysis. In these cases, the selective mixed-mode approach to the extraction of basic drugs is a suitable alternative, giving very high purity extracts with minimal levels of co-extracted material.

The ISOLUTE HCX series of mixed-mode SPE sorbents (HCX, HCX-3 and HCX-5) is based on a combination of strong cation exchange and non-polar (C8, C18 and C4 respectively) chemistries. Basic drugs are retained by two primary retention mechanisms (see **Figure 1**). This allows a rigorous interference elution regime to be used to elute interferences retained by either non-polar or cation exchange interactions alone. Only analytes with both non-polar and basic characteristics are extracted using the ISOLUTE HCX series of sorbents, providing an extremely pure final extract.



**Figure 1.** Multiple interactions on ISOLUTE HCX mixed-mode columns

The mixed-mode approach for extraction of ionizable drugs from biological fluids is extremely robust. The initial retention mechanism for the analytes is non-polar (hydrophobic), and is unaffected by the high or variable ionic strength of the matrix. The initial hydrophobic interaction is a function of the sorbent chain length, with shorter chains (e.g. C4) being less retentive than longer chains (e.g. C18). If retention of non-ionizable compounds is minimized, a cleaner extract will result.

For further information on the use of ISOLUTE HCX-3 and HCX-5, please see Technical Note **TN113**.

## **EXTRACTION PROTOCOL**

Evaluate ISOLUTE HCX, 25 mg/1 mL in the ISOLUTE Array® format using the procedure detailed below. Populate the plate with individual wells as required. Process using a VacMaster®-96 sample processing station or automated liquid handling system.

### ***Vacuum settings***

At all stages, use a short pulse (approx 1 second) of low vacuum (< -5" Hg), unless otherwise stated.

### ***Sample volume***

This procedure is optimized for a biological fluid sample volume of 100 µL. Sample should be diluted 1:1 (v/v) with appropriate buffer before applying to the column (total volume of buffered sample applied is 200 µL).

*Note:* Work in our R&D laboratory has shown that 25 mg ISOLUTE SPE columns have sufficient capacity for extraction of up to 1 mL plasma sample without analyte breakthrough. Test conditions: 1 mL plasma spiked at 0.1 mg/mL analyte concentration and diluted 1:1 with buffer before applying to the column (total volume of buffered sample applied is 2 mL).

### ***Sample pre-treatment***

Dilute the sample (100 µL of plasma or urine) with ammonium acetate buffer (0.05 M, pH 6.0, 100 µL) to give a 200 µL total sample volume at 1:1 dilution. Mix thoroughly.

### ***Column conditioning***

Place extraction plate on vacuum manifold. No collection plate should be used at this stage.

Condition each well with methanol (1 mL). Use gravity or a short pulse of vacuum to initiate flow. This will ensure efficient wetting of the hydrophobic frits, promoting even flow of sample through the wells.

### ***Column equilibration***

Rinse wells with ammonium acetate buffer (0.05 M, pH 6.0, 250 µL). Load all wells prior to applying a short pulse of vacuum to initiate flow.

### ***Sample loading***

Apply 200 µL buffered sample. Load all wells prior to applying a short pulse of vacuum to initiate flow.

### **Interference elution**

Elute acidic and neutral interferences with:

- Ammonium acetate buffer (0.05 M, pH 6.0, 250  $\mu$ L)
- Acetic acid (1 M, 250  $\mu$ L)
- Apply vacuum for 30 seconds to dry sorbent bed
- Methanol (250  $\mu$ L)

For each solvent, load all wells and allow to soak for 1 minute prior to applying a short pulse of vacuum.

### **Analyte elution**

Place collection plate in base of manifold. Ensure correct alignment (position A1 of collection plate directly underneath position A1 of extraction plate), and that extraction plate outlet Luer tips extend into the top of the collection plate. This will prevent sample cross contamination. Spacers are available to ensure optimum penetration.

Elute basic analytes with methanol/ $\text{NH}_4\text{OH}$  (95:5, v/v, 2 x 100  $\mu$ L). This will suppress ionization of the drug, overcoming both cationic and non-polar retention mechanisms, allowing elution of the analytes.

Apply the first 100  $\mu$ L aliquot and allow to soak for 2–4 mins. If the aliquot has not reached the top frit at the end of the soak time, apply a short vacuum pulse.

Apply the second 100  $\mu$ L aliquot and allow to soak for a further 2–4 mins. Apply low vacuum for 1 minute to complete elution.

Evaporate this elution solvent and re-constitute the sample in a solvent compatible with the analytical technique. For LC-MS the mobile phase is suggested.

Care should be taken to avoid losses of thermally labile or volatile analytes at this stage.

## **REAGENTS**

1. Methanol

2. 0.05 M ammonium acetate buffer pH 6

Ammonium acetate 97+% reagent, FW 77.08. Dissolve 3.854 g in 1 L of water and adjust pH using 1 M acetic acid (0.9635 g in 250 mL of water).

3. 1 M acetic acid

Acetic acid glacial 99.99+%, FW 60.05. Add 6 mL of acid to 50 mL of HPLC grade water in a 100 mL volumetric flask, make up to volume with water.

4. Methanol/ammonium hydroxide (95:5, v/v)

Add 5 mL of ammonium hydroxide, FW 35.05, to 50 mL of HPLC grade methanol in 100 mL volumetric flask, make up to volume with methanol.

## ORDERING INFORMATION

Description	Pack size	Part Number
<b>ISOLUTE Array format</b>		
ISOLUTE Array HCX 25 mg/1 mL wells*	100	902-0025-R
ISOLUTE Array HCX 25 mg/1 mL plate	1	902-0025-RP

\* As with other Array products, loose wells can be processed on a standard VacMaster-10 or -20 Sample Processing Station equipped with Array Luer adaptors (p/n 120-1201). In order to process loose wells using a VacMaster-96 Sample Processing Station, a base plate (part number 120-1000-P01) and base plate sealing strips (part number 120-1200 for sealing unused positions) are required.

Description	Pack size	Part Number
<b>ISOLUTE-96 format</b>		
ISOLUTE-96 HCX 25 mg plate	1	902-0025-P01
<b>ISOLUTE column format</b>		
ISOLUTE HCX 25 mg/1 mL	100	902-0002-A
<b>Tab-less ISOLUTE column format</b>		
ISOLUTE HCX 25 mg/1 mL (tab-less)	100	902-0002-AG

Other configurations are available, please contact Biotage for details.

### VacMaster-96 Sample Processing Station

Description	Pack size	Part Number
VacMaster-96 manifold only*	1	121-9600
VacMaster-96 Vacuum Control Unit	1	121-9601
VacMaster-96 Vacuum Control Unit with integral vacuum source	1	121-9602
VacMaster-96 with Vacuum Control Unit (121-9601)	1	121-9603
VacMaster-96 with Vacuum Control Unit (121-9602)	1	121-9604

\* Option does not include a vacuum control unit.

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